



Large-Scale Screening of the Plant Extracts for Antifungal Activity against the Plant Pathogenic Fungi

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(Received on July 21, 2022; Revised on September 27, 2022; Accepted on October 9, 2022)

Plants produce chemicals of immense diversity that provide great opportunities for development of new antifungal compounds. In search for environment-friendly alternatives to the fungicide of current use, we screened plant extracts obtained from more than eight hundred plant materials collected in Korea for their antifungal activity against the model plant pathogenic fungus, *Magnaporthe oryzae*. This initial screening identified antifungal activities from the eleven plant extract samples, among which nine showed reproducibility in the follow-up screening. These nine samples were able to suppress not only *M. oryzae* but also other fungal pathogens. Interestingly, the plant extracts obtained from *Actinostemma lobatum* comprised five out of eight samples, and were the most effective in their antifungal activity. We found that butanol fraction of the *A. lobatum* extract is the most potent. Identification and characterization of antifungal substances in the *A. lobatum* extracts would provide the promising lead compounds for new fungicide.

Keywords : antifungal activity, disease control, plant extracts

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Handling Editor : Kwang-Hyung Kim

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Over the past two decades, the incidence of plant diseases caused by fungal pathogens has dramatically increased (Fisher et al., 2012). To date, the use of agrochemicals remains one of the most effective and important agricultural practices in controlling fungal diseases of crops, although synthetic chemicals are associated with negative environmental impacts and human health concerns via direct exposure to pesticides and deposition of chemicals on agricultural products (Surapuram et al., 2014). Thus, there has been a growing need for better fungicides with less environmental burden and human health hazard. Naturally-occurring compounds are potential source of lead compounds for new agrochemicals, although being natural compounds does not necessarily guarantee being better fungicides. As plants are a cornucopia of bioactive secondary metabolites, exploiting such metabolic capacity would possibly open doors for identification and development of new disease control agents (Arif et al., 2009).

Plant extracts from many different species of plants were tested for antifungal activity against the fungal pathogens of important crops (Adnew et al., 2022; Hernández-Ceja et al., 2021; Schoss et al., 2022). In particular, some medicinal plants were heavily used for exploring possibilities of finding new lead compounds (Mohammadi et al., 2014). However, few studies attempted to survey the plant extracts sampled longitudinally from geographically diverse regions. The Nakdonggang National Institute of Biological Resources (NNIBR) has been systematically collecting and archiving diverse plant species in Korea over the years. These resources should offer opportunities to tap into the chemical repertoire of those plants.

Magnaporthe oryzae is a model plant pathogenic fungus that causes the rice blast disease. The disease is one of the most serious threats to food security in the rice-cultivating regions where the consumption of rice takes up the major

calorie intakes (Talbot, 2003). In addition to social-economic impact of the disease it causes, *M. oryzae* undergoes pathogenic development that is common to many plant pathogenic fungi, making it a model for investigating fungal development and pathogenesis. Infection usually starts with dissemination of asexual spore called conidium. The conidium that lands on leaf surface of the rice plants germinates and, upon recognition of surface hydrophobicity and hardness, differentiates an infection-specific structure, appressorium (Dean, 1997). The appressorium can build up an enormous chemical pressure that can be channeled into mechanical pressure to breach the cuticular layer of the host plants (Howard and Valent, 1996). Once inside the host cell, the fungus colonizes the plant tissues by growing inside the plant cells, leading to the formation of visible disease symptoms within a week (Kankanala et al., 2007). As the germination and subsequent development of appressorium are the critical steps that determine the onset of the disease, chemical interference with these processes is a highly effective disease control measure.

In this study, we took advantage of over eight hundred plant materials that are sampled from diverse habitats across Korea over different time points. Using collected plant materials, we initially evaluated plant extracts for the *in-vitro* antifungal activity against *M. oryzae* KJ201. With the selected extracts from the first screening, and the second screening was performed with the following important crop pathogens: *Aspergillus flavus* NRRL3357, *Botrytis cinerea*, *Rhizoctonia solani*, *Sclerotinia minor*, *Cochliobolus miyabeanus*, *Fusarium graminearum*, *Colletotrichum gloeosporioides* MGCF10, *C. gloeosporioides* YCJF8, *C. gloeosporioides* YCJF10, *Fusarium oxysporum* f. sp. *conglutinans*, and *F. oxysporum* f. sp. *niveum*. These were grown on potato dextrose agar (PDA) medium at 25°C. *M. oryzae* KJ201 and *Phytophthora infestans* T30-

4 were grown on oatmeal agar (OMA; 5% oatmeal (w/v) and 2% agar (w/v)) medium at 25°C and V8 agar medium at 16°C, respectively. For antifungal activity assays, the fungal strains were inoculated on a PDA, OMA, and V8 agar medium plate (90 mm × 15 mm Petri dish) at 25°C, 25°C and 16°C, respectively, suitable for each strain. The sterile antibiotic assay discs (6 mm diameter; Whatman, Little Chalfont, UK) were placed on the periphery of the mycelial colony, when each fungal mycelium had grown to a diameter of 20–30 mm. In addition, the 15 µl of dissolved fractionated extracts in dimethyl sulfoxide (DMSO) of 20 mg/ml were added to each antibiotic assay discs, and 15 µl of DMSO was added to the antibiotic assay disc as a negative control. After that, plates were continuously cultured at 25°C and 16°C, suitable for each strain. The growth status of each fungal mycelium was observed within 3 days.

A total of 804 plant extracts, which was obtained from the NNIBR, was first screened for their antifungal activity against the rice blast fungus (Supplementary Table 1). These 804 plant extracts were from 192 different plant species that had been collected for three years. Depending on the plant species and timing of collection, plant extracts were obtained from different tissues or organs using one of the followings: DMSO, 70% ethanol, and distilled water. For efficient screening of a large number of plant extracts, a modified disk diffusion method was used (Fig. 1A and B).

During our screens, antifungal activity was assessed not in quantitative but in binary manner (yes or no) in order to expedite the screening. The initial screening of the plant extracts led to the identification of eleven candidates. The second round of screening was performed for the eleven, and ascertained the antifungal activity for the nine candidates (Table 1). These candidates are from only five genera, and interestingly, five out of the nine candidate plant extracts were from *Actinostemma lobatum* (common name:

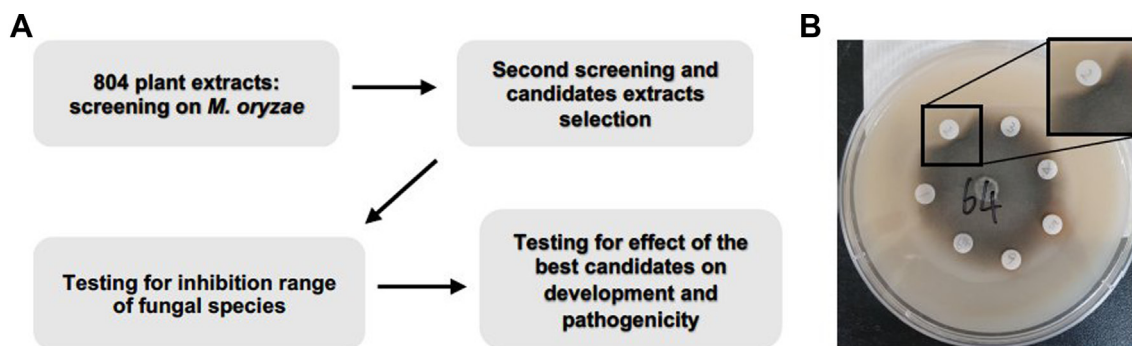


Fig. 1. Screening of plant extracts for their antifungal activity against *Magnaporthe oryzae*. (A) Flow chart for screening of plants extracts, candidate selection, and in-depth analysis. (B) Representative image of screening results based on a modified disk diffusion method. The inset figure provides magnified view of the hyphal growth inhibition by a plant extract.

Table 1. List of 11 candidate plant extracts and their antifungal activity test results in the first and second round of screening

No.	Species name	Tissue/part	Solvent	1st screen	2nd screen
1	<i>Styrax obassia</i> Siebold & Zucc.	Leaf	70% Ethanol	○	○
2	<i>Actinostemma lobatum</i> (Maxim.) Franch. & Sav.	Ground part	70% Ethanol	○	○
3	<i>Ulmus pumila</i> L.	Flower	Distilled water	○	×
4	<i>Styrax japonicus</i> Siebold & Zucc.	Leaf	Distilled water	○	○
5	<i>Actinostemma lobatum</i> (Maxim.) Franch. & Sav.	Ground part	Distilled water	○	○
6	<i>Typha angustifolia</i> L.	Leaf and stem	Distilled water	○	○
7	<i>Styrax obassia</i> Siebold & Zucc.	Leaf	Distilled water	○	○
8	<i>Phragmites japonica</i> Steud.	Above ground part	Distilled water	○	×
9	<i>Actinostemma lobatum</i> (Maxim.) Franch. & Sav.	Ground part	Distilled water	○	○
10	<i>Actinostemma lobatum</i> (Maxim.) Franch. & Sav.	Ground part	Distilled water	○	○
11	<i>Actinostemma lobatum</i> (Maxim.) Franch. & Sav.	Ground part	Distilled water	○	○

lobed *Actinostemma*), although sampling time and solvent used for extraction differ from one sample to another.

As our initial screening of the plant extracts were tested only against the single fungal pathogen, *M. oryzae*, we tested how broad the inhibitory effects of nine candidates are against the common fungal plant pathogens and an oomycete pathogen, using the modified disk diffusion method (Fig. 2A). All the extracts were effective against *S. minor* and *C. miyabeanus*. However, hyphal growth of many pathogens was inhibited only by a few plant extracts. Importantly, the plant extract #1 and #2, which correspond to 70% ethanol extracts of *S. obassia* and *A. lobatum*, respectively, showed a broad range of inhibitory effect against the pathogens tested here (Table 1, Fig. 2A).

Based on the abundance of samples and broad range of inhibitory effect, we prioritized the *A. lobatum* for further analysis using different solvents. *A. lobatum*, an annual herb of Cucurbitaceae family, is widespread in Asia (Kim, 2010; Li et al., 2012) and it has traditionally been used as a medicinal herb for the treatment of arthritis, cardiovascular disease and detoxification of snake bite (Cao et al., 2015; Kim et al., 2008; Li et al., 2012). Although biological activities of *A. lobatum* such as anti-tumor activity (Fujioka et al., 1996; Li et al., 2012), anti-thrombotic activity (Kim et al., 2008), anti-oxidative activity (Kim, 2010), and anti-bacterial activity (Chen et al., 2005) have been reported, its anti-fungicidal activity has not been investigated to date. Dried and powdered *A. lobatum* (3.3 kg) was extracted successively with 70% ethanol (2 × 10 l) at room temperature and filtered. The filtrate was evaporated under a vacuum to obtain a 70% EtOH extract. The crude extract (390 g, yield to 11.8%) was re-suspended in water and partitioned successively with hexane (107.7 g), chloroform (11.6 g), ethyl acetate (21.1 g), and normal butanol (60.1 g).

When we tested the inhibitory effect of individual extracts obtained by different solvents on a number of common plant pathogens (Fig. 2B), the butanol extract showed the strongest inhibitory effect on almost all the pathogens tested, suggesting the presence of active compound(s) in the butanol fraction. To gauge the potency of butanol fraction, we compared the butanol fraction with hygromycin B, which is a commonly used aminoglycoside antibiotic produced by *Streptomyces hygroscopicus* (Eustice and Wilhelm, 1984; González et al., 1978; Mann and Bromer, 1958). Such comparison showed that butanol fraction is able to inhibit hyphal growth of the fungus more effectively than the equivalent amount of hygromycin (Supplementary Fig. 1A). We also compared the antifungal activity of butanol extract with that of a fungicide, benomyl. This indicated that butanol extract has weaker antifungal activity than benomyl in our plate assay. These results suggest that butanol fraction of *A. lobatum* contains compound(s) having higher potency than hygromycin B.

Since our analyses focused only on inhibition of hyphal growth by the plant extracts, we next tested the effects of the butanol fraction on pathogenic development of the rice blast fungus. The *M. oryzae* KJ201 strain cultured on OMA medium at 25°C under constant light to promote conidium production. Conidia were harvested from 9-day-old colonies using 5 ml of sterilized distilled water and counted using a hemocytometer under a light microscope. To monitor germination and appressorium formation, the concentration of conidial suspension was adjusted to $\sim 5 \times 10^4$ conidia/ml. A 40 μ l suspension was placed on a hydrophobic surface of plastic coverslips and then incubated in a humidity chamber at 25°C. The germination and appressorium formation rate were calculated by counting the number of germinating spores and appressorium-forming conidia, respectively, out

A

Pathogen species	Plant extracts									
	1	2	3	5	6	7	8	10	11	
<i>A. flavus</i>										
<i>B. cinerea</i>										
<i>R. solani</i>										
<i>S. minor</i>										
<i>C. miyabeanus</i>										
<i>F. graminearum</i>										
<i>C. gloeosporioides</i> MGCF10										
<i>F. oxysporum</i> f.sp. <i>conglutinans</i>										
<i>P. infestans</i>										

B

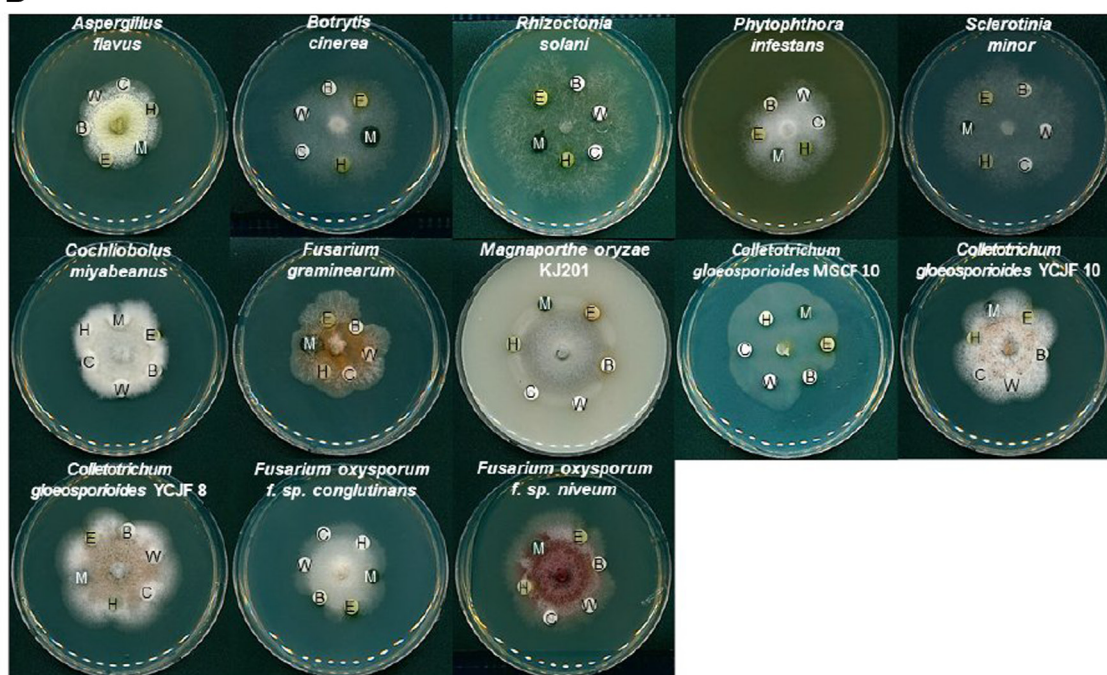


Fig. 2. Antifungal activity of selected plant extracts against a set of common plant pathogens. (A) A heatmap summarizing antifungal activity test results against a set of common plant pathogens. Results were shown as binary outcome: inhibition, gray-filled block; no inhibition, white-block. *A. flavus*, *Aspergillus flavus*; *B. cinerea*, *Botrytis cinerea*; *R. solani*, *Rhizoctonia solani*; *S. minor*, *Sclerotinia minor*; *C. miyabeanus*, *Cochliobolus miyabeanus*; *F. graminearum*, *Fusarium graminearum*; *C. gloeosporioides*, *Colletotrichum gloeosporioides*; *F. oxysporum*, *Fusarium oxysporum*; *P. infestans*, *Phytophthora infestans*. (B) Effects of solvents used in extraction on antifungal activity of *Actinostemma lobatum* extract. C, dimethyl sulfoxide; B, *A. lobatum* butanol extract; E, *A. lobatum* ethyl acetate extract; H, *A. lobatum* hexane extract; M, *A. lobatum* chloroform extract; W, *A. lobatum* water extract.

of at least 100 spores at 2, 4, and 6 h post-incubation. All the assays were conducted with three technical replicates in three independent experiments. Slides observed under a DM2500 microscope (Leica, Wetzlar, Germany) and pictured with a DFC7000 T digital camera (Leica) using LAS X software (Leica).

The harvested spores were incubated with varying con-

centrations of the butanol fraction on inductive surface (plastic coverslip) (Fig. 3A). In contrast to the DMSO control in which spores germinate and develop appressoria over time, a high concentration (300 $\mu\text{g/ml}$ and 150 $\mu\text{g/ml}$) of the butanol fraction completely inhibited the spore germination itself (Fig. 3A and B). At a much lower concentration (30 $\mu\text{g/ml}$), although inhibition of spore germination

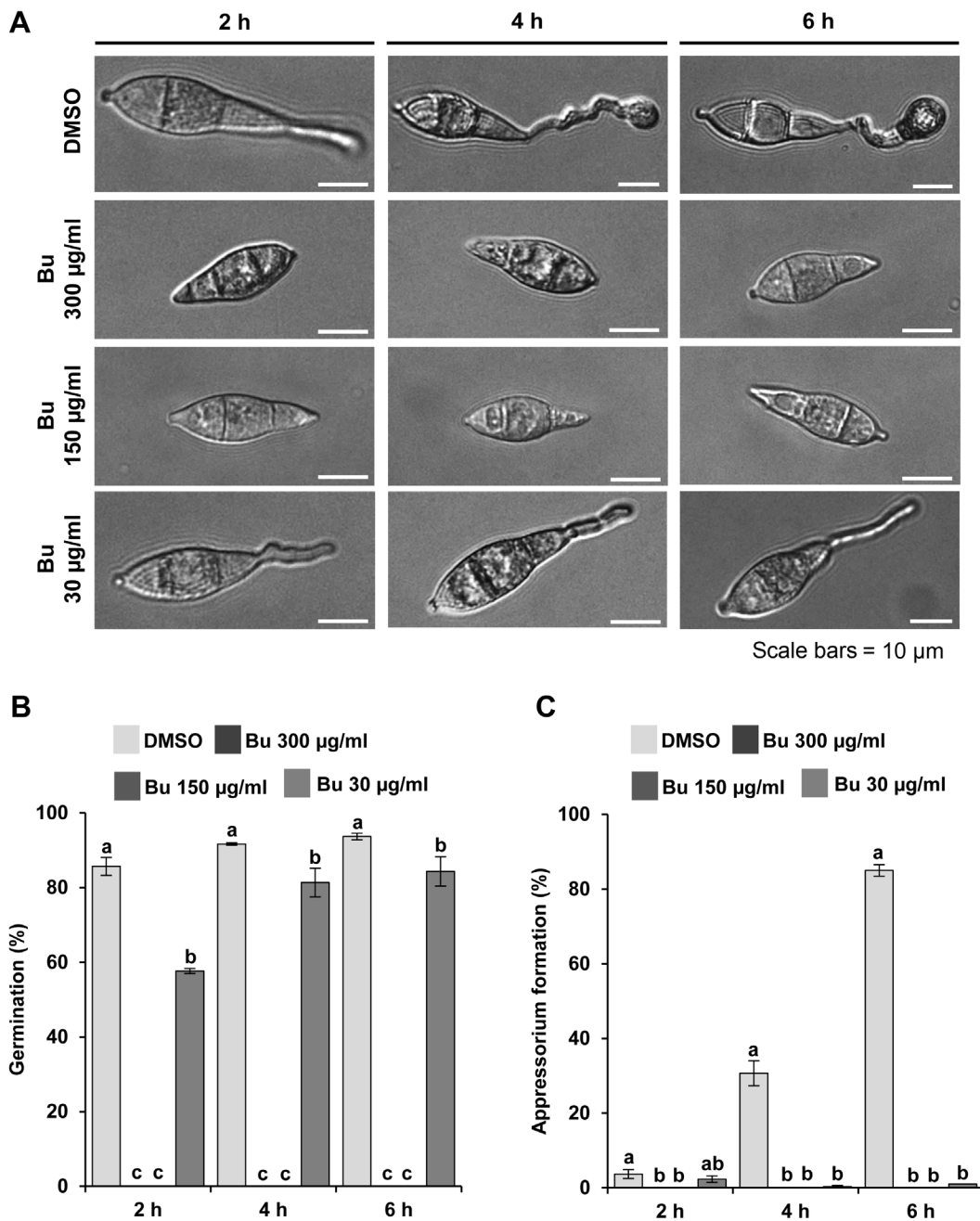


Fig. 3. Effects of *Actinostemma lobatum* butanol extracts (Bu) on fungal development. (A) Images showing the effects of varying concentration of Bu on spore germination and appressorium of *Magnaporthe oryzae* KJ201 at 2, 4, and 6 h post-incubation of spores on hydrophobic surface. (B) Quantitative measurement of germination under Bu treatment. (C) Quantitative measurement of appressorium formation rates under Bu treatment. Different letters on the bars indicate significant difference in mean values ($P < 0.001$, Tukey honestly significant difference test, $n = 3$).

was significantly alleviated, appressorium formation was rarely observed, indicating that the butanol fraction has inhibitory effect on fungal development as well (Fig. 3A-C).

Furthermore, pathogenicity was carried out to test whether the extract is effective in suppressing disease de-

velopment. For pathogenicity assays, rice plants (*Oryza sativa* cv. Dongjin) were used in three- to four-leaf stages grown at 25°C for 4 weeks. To carry out wound inoculation, 40 µl of spore suspension ($\sim 5 \times 10^5$ spores/ml) was directly placed onto wounded site of rice leaves. Inoculated

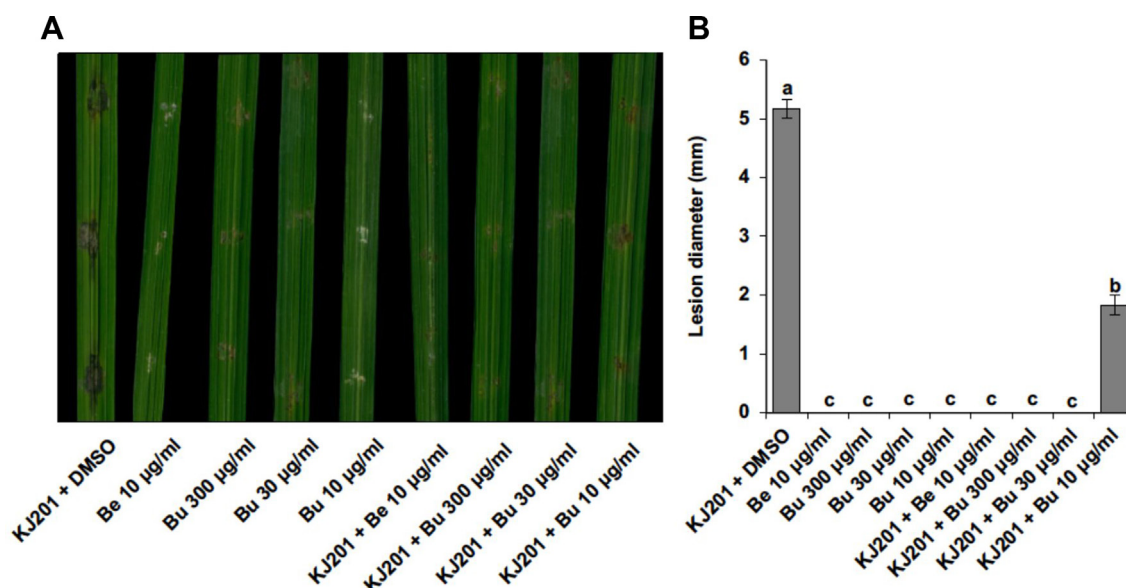


Fig. 4. Effects of *Actinostemma lobatum* butanol extracts (Bu) on fungal pathogenicity. (A) Assessment of pathogenicity of *Magnaporthe oryzae* KJ201 using wound inoculation with or without the Bu. (B) Quantitative analysis of pathogenicity assay results obtained by measurement of lesion length. Be, benomyl. Different letters on the bars indicate statistically significant difference in mean values ($P < 0.001$, Tukey honestly significant difference test, $n = 3$).

plants were incubated in a humidity chamber without light for 24 h and then transferred to an incubator for growth at 25°C with a 16 h photoperiod of fluorescent light. After 7 d post-inoculation, lesion development was assessed and photographed. Plants grown in three pots were used for a pathogenicity assay in three independent experiments. Our pathogenicity assay using detached rice leaves showed that butanol fraction is able to significantly suppress the disease symptom development with little side-effect on the plants even at higher concentration (Fig. 4A and B). Importantly, the butanol extract showed disease control activity that is almost comparable to benomyl (Fig. 4A and B).

In this work, we have screened a large number of plant materials covering diverse geographical locations and species in Korea in order to identify the extracts having antifungal activities. This effort has shortlisted a few plant species including *A. lobatum* as potential candidates for developing plant-derived agrochemicals. Molecular identification and characterization of active compound(s) in the butanol fraction might yield a novel agrochemical(s) in the years to come.

Conflicts of Interest

No potential conflict of interest relevant to this article was reported.

Acknowledgments

This work was supported by a grant from the Nakdong-gang National Institute of Biological Resources (NNIBR), funded by the Ministry of Environment (MOE) of the Republic of Korea (NNIBR202202110), and by a grant from the National Research Foundation of Korea (NRF-2018R1A5A1023599).

Electronic Supplementary Material

Supplementary materials are available at The Plant Pathology Journal website (<http://www.ppjonline.org/>).

References

- Adnew, A., Shifa, H. and Mohammed, A. 2022. Antifungal activity of plant extracts against postharvest mould fungi associated with coffee (*Coffea arabica* L.) in Bale Zone, Ethiopia. *Org. Agric.* 12:107-124.
- Arif, T., Bhosale, J. D., Kumar, N., Mandal, T. K., Bendre, R. S., Lavekar, G. S. and Dabur, R. 2009. Natural products--antifungal agents derived from plants. *J. Asian Nat. Prod. Res.* 11:621-638.
- Cao, J.-Q., Li, W., Tang, Y., Zhang, X.-r., Li, W. and Zhao, Y.-Q. 2015. Three new triterpene saponins from *Actinostemma lobatum* Maxim. and their cytotoxicity *in vitro*. *Phytochem.*

- Lett.* 11:301-305.
- Chen, Y., Lin, R., Yang, S. and Gao, Z. 2005. Study on the antibacterial activity of *Actinostemma tenerum* polysaccharides. *Prevent. Med. Tribune* 15:1240-1241.
- Dean, R. A. 1997. Signal pathways and appressorium morphogenesis. *Annu. Rev. Phytopathol.* 35:211-234.
- Eustice, D. C. and Wilhelm, J. M. 1984. Mechanisms of action of aminoglycoside antibiotics in eucaryotic protein synthesis. *Antimicrob. Agents Chemother.* 26:53-60.
- Fisher, M. C., Henk, D. A., Briggs, C. J., Brownstein, J. S., Madoff, L. C., McCraw, S. L. and Gurr, S. J. 2012. Emerging fungal threats to animal, plant and ecosystem health. *Nature* 484:186-194.
- Fujioka, T., Kashiwada, Y., Okabe, H., Mihashi, K. and Lee, K.-H. 1996. Antitumor agents 171. Cytotoxicities of lobatosides B, C, D, and E, cyclic bisdesmosides isolated from *Actinostemma lobatum* Maxim. *Bioorg. Med. Chem. Lett.* 6:2807-2810.
- González, A., Jiménez, A., Vázquez, D., Davies, J. E. and Schindler, D. 1978. Studies on the mode of action of hygromycin B, an inhibitor of translocation in eukaryotes. *Biochim. Biophys. Acta* 521:459-469.
- Hernández-Ceja, A., Loeza-Lara, P. D., Espinosa-García, F. J., García-Rodríguez, Y. M., Medina-Medrano, J. R., Gutiérrez-Hernández, G. F. and Ceja-Torres, L. F. 2021. *In vitro* antifungal activity of plant extracts on pathogenic fungi of blueberry (*Vaccinium* sp.). *Plants (Basel)* 10:852.
- Howard, R. J. and Valent, B. 1996. Breaking and entering: host penetration by the fungal rice blast pathogen *Magnaporthe grisea*. *Annu. Rev. Microbiol.* 50:491-512.
- Kankanala, P., Czymbek, K. and Valent, B. 2007. Roles for rice membrane dynamics and plasmodesmata during biotrophic invasion by the blast fungus. *Plant Cell* 19:706-724.
- Kim, D. K. 2010. Antioxidative constituents from the whole plant of *Actinostemma lobatum* Maxim. *J. Korean Soc. Appl. Biol. Chem.* 53:746-751.
- Kim, K.-H., Lee, H.-J., Lee, J.-H., Jang, Y.-S., Kim, D.-K., Shim, B.-S., Cho, K.-H., Ko, S.-G., Ahn, K.-S. and Kim, S.-H. 2008. Blockade of glycoprotein IIb/IIIa mediates the antithrombotic activity of butanol fraction of *Actinostemma lobatum* Maxim. *J. Ethnopharmacol.* 116:431-438.
- Li, W., Cao, J., Tang, Y., Zhang, L., Xie, Q., Shen, H. and Zhao, Y. 2012. Cyclic bisdesmosides from *Actinostemma lobatum* MAXIM (Cucurbitaceae) and their *in vitro* cytotoxicity. *Fito-terapia* 83:147-152.
- Mann, R. L. and Bromer, W. W. 1958. The isolation of a second antibiotic from *Streptomyces hygroscopicus*. *J. Am. Chem. Soc.* 80:2714-2716.
- Mohammadi, A., Nazari, H., Imani, S. and Amrollahi, H. 2014. Antifungal activities and chemical composition of some medicinal plants. *J. Mycol. Med.* 24:e1-e8.
- Schoss, K., Kočevar Glavač, N., Dolenc Koče, J. and Anžlovar, S. 2022. Supercritical CO₂ plant extracts show antifungal activities against crop-borne fungi. *Molecules* 27:1132.
- Surapuram, V., Setzer, W. N., McFeeters, R. L. and McFeeters, H. 2014. Antifungal activity of plant extracts against *Aspergillus niger* and *Rhizopus stolonifer*. *Nat. Prod. Commun.* 9:1603-1605.
- Talbot, N. J. 2003. On the trail of a cereal killer: exploring the biology of *Magnaporthe grisea*. *Annu. Rev. Microbiol.* 57:177-202.